

Psych: A Swiss Army Knife for psychology

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Outline

A Swiss Army knife for psychologists

Preliminaries

Data entry and description

Getting and cleaning data

Graphical displays

Multivariate analysis

The number of factors problem

factors and clusters

Hierarchical models

True hierarchical

Seemingly hierarchical

Scale Construction

From raw data

From correlation matrices

The many forms of reliability

Regression, moderation, mediation

Dust bowl empiricism

The psych and psychTools packages

1. *psych* and *psychTools* have been developed to help further research in personality and individual differences.
 2. Like a Swiss Army Knife, it is not the best tool for anything, but it is a very helpful tool for many things.
 3. It is particularly aimed at the researcher in personality and individual differences.
 4. Like core R. it helps researchers do open source science.
 5. It is meant to be a relatively “light” package, in that it does not have many dependencies.
 6. Unlike many other packages, the Help pages and Vignettes are fairly extensive (some would say wordy).

Installing the psych package (\geq 2.3.6)

R code

```
#if you have not already done so, you first install the package  
install.packages("psych",dependencies=TRUE)  
  
library(psych) #you need to do this every time you start R  
  
#or automate the library(psych) call  
#by creating and saving a function  
  
.First <- function() {library(psych)}  
quit() #with save option  
  
#start R and psych will be automatically loaded  
sessionInfo() #will tell you what version you are using
```

Good morning Bill.

Are you ready to have some fun?

```
> sessionInfo()
version 4.3.1 (2023-06-16)
Platform: aarch64-apple-darwin20 (64-bit)
Running under: macOS Ventura 13.4.1
```

To see the dependencies

R code

`sessionInfo()`

```
sessionInfo()
R version 4.3.1 (2023-06-16)
Platform: aarch64-apple-darwin20 (64-bit)
Running under: macOS Ventura 13.4.1

Matrix products: default
BLAS:      /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRblas.0.dylib
LAPACK:   /Library/Frameworks/R.framework/Versions/4.3-arm64/Resources/lib/libRlapack.dylib

Random number generation:
RNG:      Mersenne-Twister
Normal:   Inversion
Sample:   Rounding

locale:
[1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
time zone: America/Chicago
tzcode source: internal

attached base packages:
[1] stats      graphics   grDevices utils       datasets   methods    base

other attached packages:
[1] psychTools_2.3.6 psych_2.3.6

loaded via a namespace (and not attached):
[1] compiler_4.3.1 tools_4.3.1    parallel_4.3.1 foreign_0.8-84 nlme_3.1-162 mnormt_2.6.178
```

Show all the functions in the psych package

```
objects("package:psych")
```

```
[1] "%+%"          "acs"           "alpha"          "alpha.ci"
[5] "alpha2r"       "anova.psych"   "AUC"            "autoR"
[9] "bassAckward"  "bassAckward.diagram" "Bechtoldt"      "Bechtoldt.ci"
[13] "Bechtoldt.2"  "bestItems"      "bestScales"     "bf1"
[17] "bfi.keys"     "bi.bars"        "bifactor"       "bigCor"
[21] "biplot.psych" "biquartimin"    "biserial"        "block.rando"
[25] "bock.table"    "cattell"         "cd.validity"   "char2numer"
[29] "Chen"          "chi2r"          "circ.sim"       "circadian.cor"
[33] "circ.simulation" "circ.tests"     "circadian.mean" "circadian.phase"
[37] "circadian.linear.cor" "circadian.stats" "circadian.stats" "circadian.cor"
[41] "circadian.sd"  "cluster.cor"    "circular.cor"  "circadian.phase"
[45] "cluster.cor"   "cluster.fit"    "cluster.loadings" "circular.me"
[49] "cluster2keys"  "cohen.d"        "cohen.d.by"    "cluster.plot"
[53] "cohen.kappa"   "cohen.profile" "comorbidity"   "cohen.d.ci"
[57] "congeneric.sim" "congruence"    "cor.ci"         "con2cat"
[...]
[441] "superMatrix"   "t2d"            "t2r"            "cor.plot"
[445] "table2matrix"  "tableF"         "Tal.Or"         "table2df"
[449] "target.rot"    "TargetQ"        "TargetT"        "Tal.Or"
[453] "test.all"      "test.irt"        "test.psych"    "tenberge"
[457] "tetrachoric"   "thurstone"      "Thurstone"      "testRetest"
[461] "Thurstone.33G" "Thurstone.9"   "topBottom"      "Thurstone.ci"
[465] "Tucker"         "unidim"         "validityItem"  "tr"
[469] "vgQ.bimin"     "vgQ.targetQ"  "vgQ.varimin"   "varimin"
[473] "violinBy"       "vss"             "VSS"            "violin"
[477] "VSS.plot"       "VSS.scree"       "VSS.sim"        "VSS.parallel"
[481] "West"            "winsor"          "winsor.mean"   "VSS.simulation"
[485] "winsor.sd"      "winsor.var"      "withinBetween" "winsor.mean"
[489] "Yule"            "Yule.inv"        "Yule2phi"      "wkappa"
[493] "Yule2poly"      "Yule2poly.matrix" "Yule2tetra"    "Yule2phi.matrix"
[497] "YuleCor"
```

Objects in *psychTools*

```
objects(package:psychTools)
[1] "ability"
[4] "all.income"
[7] "Athenstaedt.keys"
[10] "bfi.adjectives.keys"
[13] "big5.100.adjectives"
[16] "blot"
[19] "city.location"
[22] "colom.ed1"
[25] "combineMatrices"
[28] "cushny"
[31] "dfOrder"
[34] "epi.bfi"
[37] "epiR"
[40] "fileScan"
[43] "galton"
[46] "GERAS.keys"
[49] "heights"
[52] "holzinger.swineford"
[55] "iqitems"
[58] "msq.keys"
[61] "omega2latex"
[64] "read.clipboard"
[67] "read.clipboard.lower"
[70] "read.file"
[73] "recode"
[76] "sat.act"
[79] "Spengler"
[82] "spi.dictionary"
[85] "tai"
[88] "vJoin"
[91] "zola"

[1] "ability.keys"
[4] "Athenstaedt"
[7] "bfi"
[10] "bfi.dictionary"
[13] "big5.adjectives.keys"
[16] "burt"
[19] "colom"
[22] "colom.ed2"
[25] "cor2latex"
[28] "Damian"
[31] "eminence"
[34] "epi.dictionary"
[37] "fa2latex"
[40] "filesInfo"
[43] "GERAS.dictionary"
[46] "GERAS.scales"
[49] "holzinger.dictionary"
[52] "ICC2latex"
[55] "irt2latex"
[58] "msqR"
[61] "peas"
[64] "read.clipboard.csv"
[67] "read.clipboard.tab"
[70] "read.file.csv"
[73] "sai"
[76] "Schutz"
[79] "Spengler.stat"
[82] "spi.keys"
[85] "USAF"
[88] "write.file"
[91] "zola.dictionary"

[1] "affect"
[4] "Athenstaedt.dictionary"
[7] "bfi.adjectives.dictionary"
[10] "bfi.keys"
[13] "blant"
[16] "cities"
[19] "colom.ed0"
[22] "colom.ed3"
[25] "cubits"
[28] "df2latex"
[31] "epi"
[34] "epi.keys"
[37] "fileCreate"
[40] "filesList"
[43] "GERAS.items"
[46] "globalWarm"
[49] "holzinger.raw"
[52] "income"
[55] "msq"
[58] "neo"
[61] "Pollack"
[64] "read.clipboard.fwf"
[67] "read.clipboard.upper"
[70] "read.https"
[73] "sai.dictionary"
[76] "selectBy"
[79] "spi"
[82] "splitBy"
[85] "veg"
[88] "write.file.csv"
[91] "zola.keys"
```

Vignettes and How To's

- How to's and Vignettes
 1. An introduction to the *psych* package: Part I:.
 2. Intro:Part II: Scale construction and psychometrics
 3. Installing R and the *psych* package
 4. Using R and *psych* to find ω
 5. How To: Use *psych* for factor analysis and data reduction
 6. Using R to score personality scales
 7. Using *psych* for regression and mediation analysis
 - User manual for *psych*
 - User manual for *psychTools*
 - Help files for *psych*
 - Help files for *psychTools*

Get your data: using `read.file` or `read.clipboard`

From a website: define the file name

frame R code

```
fn <- "https://personality-project.org/r/datasets/glbwarm.sav"
fn #show it to check
fn
[1] "https://personality-project.org/r/datasets/glbwarm.sav"
mydata <- read.table(fn)
```

From a local file: find the file using `read_file`

String code

```
> my.data <- read.file() #will open a search window, read the file  
#depending upon the suffix, will read .sav, .csv, .txt,  
#.rds, .rDa, etc.
```

From the clipboard: (first, go to the remote site, copy to the clipboard and then use the `read.clipboard` function).

R code

```
mydata <- read.clipboard()      #or  
mydata <- read.clipboard.tab()  #if an excel file  
my.data <- read.clipboard.csv() #if a tab delimited file
```

(This example data set can also be accessed directly in `glbwarm`.)

R code

```
dim(mydata) #how many rows and columns?  
headTail(mydata) #Show the top and bottom n rows and columns from cl  
describe(mydata) #basic descriptive statistics
```

```

dim(mydata) #how many rows and columns?
[1] 815    7
> headTail(mydata) #Show the top and bottom n rows and columns from c1 to c2
   govact posemot negemot ideology age sex partyid
1     3.6    3.67    4.67      6  61    0       2
2      5        2    2.33      2  55    0       1
3     6.6    2.33    3.67      1  85    1       1
4      1        5        5      1  59    0       1
...
812    3.4        1        1      7  67    0       3
813    1.6    3.67    1.67      7  72    1       3
814    5.4    2.67    3.33      6  36    0       2
815    5.4    5.33        6      4  82    1       1
> describe(mydata) #basic descriptive statistics
   vars   n   mean     sd median trimmed    mad min max range skew kurtosis   se
govact    1 815  4.59  1.36    4.80   4.68  1.19    1    7    6 -0.63  0.22 0.05
posemot    2 815  3.13  1.35    3.00   3.11  1.48    1    6    5  0.09 -0.85 0.05
negemot    3 815  3.56  1.53    3.67   3.58  1.97    1    6    5 -0.15 -1.07 0.05
ideology   4 815  4.08  1.51    4.00   4.07  1.48    1    7    6  0.03 -0.43 0.05
age        5 815 49.54 16.33   51.00  49.66 19.27   17   87   70 -0.07 -1.03 0.57
sex        6 815  0.49  0.50    0.00   0.49  0.00    0    1    1  0.05 -2.00 0.02
partyid   7 815  1.88  0.87    2.00   1.85  1.48    1    3    2  0.23 -1.63 0.03
>

```

headTail of a bigger local file

R code

```
dim(msqR)
headTail(msqR,top=4,bottom=6,from=78, to=88)
```

```
[1] 6411 88
headTail(msqR,top=4,bottom=6,from=78, to=88)
   Lie Sociability Impulsivity gender TOD drug film time id form study
1    3         7          1     2   9   2 <NA>    1   1   2 AGES
2    3         9          4     2   9   1 <NA>    1   2   2 AGES
3    4         3          1     1   9   1 <NA>    1   3   2 AGES
4    1        11          4     2   9   2 <NA>    1   4   2 AGES
...
3941  2         9          6 <NA>    9   2   4    2 195   2 XRAY
3942  3         3          5 <NA>    9   1   2    2 196   2 XRAY
3943  2         7          2 <NA>    9   2   1    2 197   2 XRAY
3944  0        12          5 <NA>    9   1   4    2 198   2 XRAY
3945  3        11          3 <NA>    9   1   3    2 199   2 XRAY
3946  0         2          4 <NA>    9   2   4    2 200   2 XRAY
```

Notice that rowname although unique is not the case Number

Descriptives by a grouping variable

R code

```
describeBy(mydata~sex)
```

Descriptive statistics by group

sex: 0

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis	se
govact	1	417	4.72	1.16	4.8	4.77	1.19	1	7	6	-0.52	0.55	0.06
posemot	2	417	3.03	1.39	3.0	3.00	1.48	1	6	5	0.17	-0.98	0.07
negemot	3	417	3.73	1.45	4.0	3.79	1.48	1	6	5	-0.26	-0.83	0.07
ideology	4	417	3.89	1.44	4.0	3.87	1.48	1	7	6	0.05	-0.29	0.07
age	5	417	46.90	14.95	44.0	46.77	16.31	18	83	65	0.13	-0.90	0.73
sex	6	417	0.00	0.00	0.0	0.00	0.00	0	0	0	NaN	NaN	0.00
partyid	7	417	1.79	0.86	2.0	1.74	1.48	1	3	2	0.41	-1.52	0.04

sex: 1

	vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis	se
govact	1	398	4.45	1.53	4.60	4.55	1.48	1	7	6	-0.56	-0.31	0.08
posemot	2	398	3.23	1.30	3.33	3.23	1.48	1	6	5	0.04	-0.68	0.07
negemot	3	398	3.37	1.59	3.67	3.36	1.98	1	6	5	-0.01	-1.25	0.08
ideology	4	398	4.29	1.56	4.00	4.29	1.48	1	7	6	-0.05	-0.56	0.08
age	5	398	52.30	17.25	55.00	52.85	19.27	17	87	70	-0.33	-1.01	0.86
sex	6	398	1.00	0.00	1.00	1.00	0.00	1	1	0	NaN	NaN	0.00
partyid	7	398	1.98	0.87	2.00	1.98	1.48	1	3	2	0.04	-1.67	0.04

Correlations using `lowerCor`

`lowerCor` is a nice example of the power of R to nest functions. It is just a call to `cor` with the `use="pairwise"` option followed by a call to `lowerMat` which “prettifies” a correlation matrix.

R code

```
R<- lowerCor(mydata)    #returns R invisibly
```

```

lowerCor(mydata)
            govct posmt negmt idlgy age     sex    prtyd
govact      1.00
posemot      0.04  1.00
negemot      0.58  0.13  1.00
ideology   -0.42 -0.03 -0.35  1.00
age        -0.10  0.04 -0.06  0.21  1.00
sex        -0.10  0.07 -0.12  0.13  0.17  1.00
partvid   -0.36 -0.04 -0.32  0.62  0.15  0.11  1.00

```

R	#show R (if you want to use it for something else. Note that it is not rounded.	govact	posemot	negemot	ideology	age	sex	partyid
govact	1.00000000	0.04302895	0.57774582	-0.41831995	-0.09713873	-0.09861854	-0.36039647	
posemot	0.04302895	1.00000000	0.12792202	-0.02937618	0.04235193	0.07429449	-0.03577099	
negemot	0.57774582	0.12792202	1.00000000	-0.34878643	-0.05689493	-0.11735643	-0.32419141	
ideology	-0.41831995	-0.02937618	-0.34878643	1.00000000	0.21240565	0.13288895	0.61945381	
age	-0.09713873	0.04235193	-0.05689493	0.21240565	1.00000000	0.16553039	0.15443184	
sex	-0.09861854	0.07429449	-0.11735643	0.13288895	0.16553039	1.00000000	0.10875960	
partyid	-0.36039647	-0.03577099	-0.32419141	0.61945381	0.15443184	0.10875960	1.00000000	

Correlations using corr.test

R code

```
corr.test(mydata)
```

```

Call:corr.test(x = mydata)
Correlation matrix
      govact posemot negemot ideology   age   sex partyid
govact    1.00    0.04    0.58   -0.42 -0.10 -0.10   -0.36
posemot   0.04    1.00    0.13   -0.03  0.04  0.07   -0.04
negemot   0.58    0.13    1.00   -0.35 -0.06 -0.12   -0.32
ideology -0.42   -0.03   -0.35    1.00  0.21  0.13    0.62
age       -0.10   0.04   -0.06    0.21  1.00  0.17    0.15
sex       -0.10   0.07   -0.12    0.13  0.17  1.00    0.11
partyid  -0.36   -0.04   -0.32    0.62  0.15  0.11    1.00
Sample Size
[1] 815
Probability values (Entries above the diagonal are adjusted* for multiple tests.)
      govact posemot negemot ideology   age   sex partyid
govact    0.00    0.88    0.0    0.00  0.04  0.04    0.00
posemot   0.22    0.00    0.0    0.88  0.88  0.20    0.88
negemot   0.00    0.00    0.0    0.00  0.52  0.01    0.00
ideology  0.00    0.40    0.0    0.00  0.00  0.00    0.00
age       0.01    0.23    0.1    0.00  0.00  0.00    0.00
sex       0.00    0.03    0.0    0.00  0.00  0.00    0.02
partyid  0.00    0.31    0.0    0.00  0.00  0.00    0.00

```

To see confidence intervals of the correlations, print with the short=FALSE option:

*Adjustment using the Holm (1979) correction for multiple tests

long output from corr.test gives the normal theory CI

R code

```
print(corr.test(mydata), short=FALSE)
```

	Confidence intervals based upon normal theory. To get bootstrapped values, try cor.ci					
	raw.lower	raw.r	raw.upper	raw.p	lower.adj	upper.adj
govct-posmt	-0.03	0.04	0.11	0.22	-0.04	0.13
govct-negmt	0.53	0.58	0.62	0.00	0.50	0.64
govct-idlgy	-0.47	-0.42	-0.36	0.00	-0.50	-0.33
govct-age	-0.16	-0.10	-0.03	0.01	-0.19	0.00
govct-sex	-0.17	-0.10	-0.03	0.00	-0.19	0.00
govct-prtyd	-0.42	-0.36	-0.30	0.00	-0.45	-0.27
posmt-negmt	0.06	0.13	0.19	0.00	0.03	0.22
posmt-idlgy	-0.10	-0.03	0.04	0.40	-0.10	0.04
posmt-age	-0.03	0.04	0.11	0.23	-0.04	0.13
posmt-sex	0.01	0.07	0.14	0.03	-0.02	0.17
posmt-prtyd	-0.10	-0.04	0.03	0.31	-0.11	0.04
negmt-idlgy	-0.41	-0.35	-0.29	0.00	-0.44	-0.25
negmt-age	-0.13	-0.06	0.01	0.10	-0.15	0.03
negmt-sex	-0.18	-0.12	-0.05	0.00	-0.21	-0.02
negmt-prtyd	-0.38	-0.32	-0.26	0.00	-0.41	-0.23
idlgy-age	0.15	0.21	0.28	0.00	0.11	0.31
idlgy-sex	0.06	0.13	0.20	0.00	0.03	0.23
idlgy-prtyd	0.58	0.62	0.66	0.00	0.55	0.68
age-sex	0.10	0.17	0.23	0.00	0.06	0.26
age-prtyd	0.09	0.15	0.22	0.00	0.05	0.25
sex-prtyd	0.04	0.11	0.18	0.00	0.01	0.20

Adjusted cis are given with Holm (1979) adjustment

Correlations with “magic asterisks”

R code

```
print(corr.test(mydata)$stars, quote=FALSE)
```

```

print(corr.test(mydata)$stars, quote=FALSE)
      govact posemot negemot ideology age sex partyid
govact   1***    0.04   0.58*** -0.42*** -0.1* -0.1* -0.36*** 
posemot   0.04   1***   0.13** -0.03    0.04   0.07  -0.04
negemot   0.58***  0.13*** 1***   -0.35*** -0.06  -0.12** -0.32*** 
ideology -0.42*** -0.03   -0.35*** 1***   0.21*** 0.13** 0.62*** 
age       -0.1**  0.04   -0.06   0.21*** 1***   0.17*** 0.15*** 
sex       -0.1**  0.07*  -0.12*** 0.13*** 0.17*** 1***   0.11* 
partyid  -0.36*** -0.04   -0.32*** 0.62*** 0.15*** 0.11** 1*** 

```

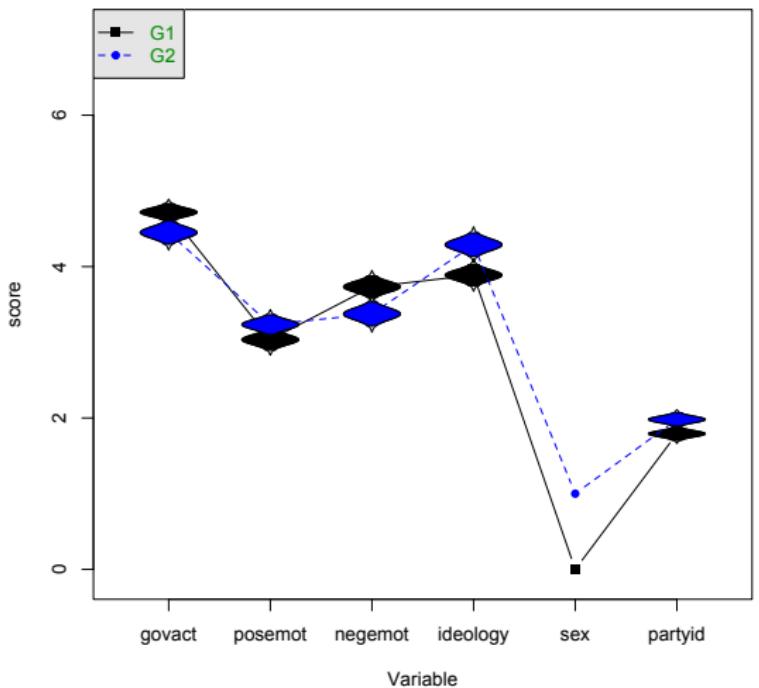
Once again, the p values above the diagonal are adjusted using the [Holm \(1979\)](#) correction for multiple tests.

Graphical displays of data

1. Can show basic means and ranges (error.bars and error.bars.by)
 2. Can show correlations using pairs.panels, corPlot
 3. densityBy for distribution information.
 4. scatterHist to show bivariate plots with densities by group
 5. cohen.d combines with error.dots to show effect sizes and confidence intervals

Showing group differences using error.bars.by

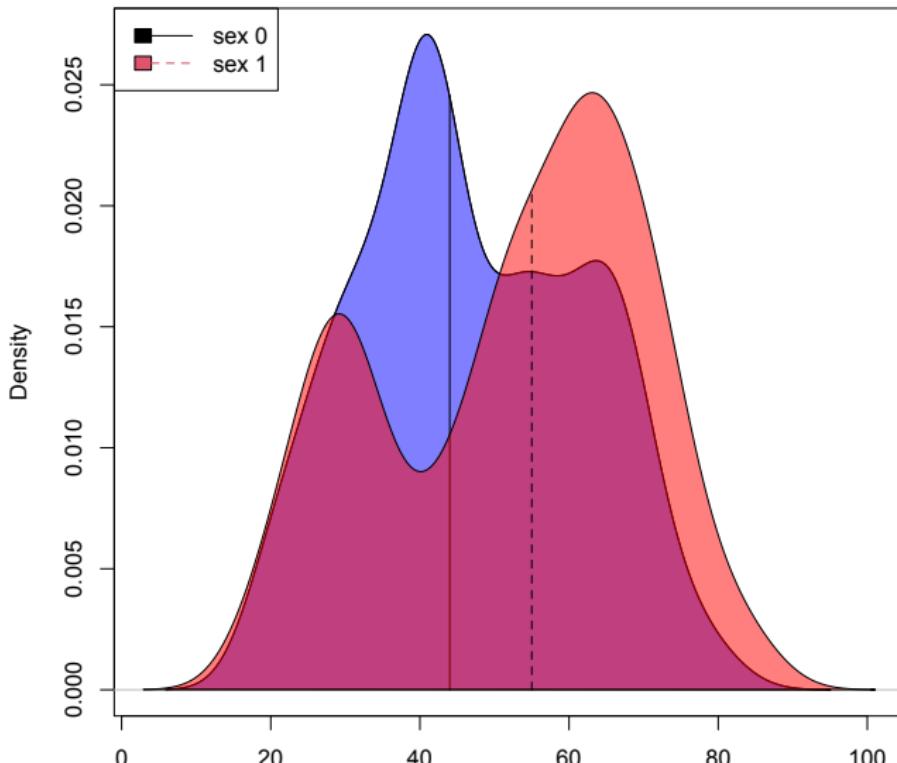
95% confidence limits



```
error.bars.by(mydata[-5], "sex", by.var=FALSE, ylab="score", xlab="Variable")
```

Global warming data set – age by sex

Age distributions for Global Warming data



Finding the effect of on mood

1. What is the effect of caffeine on motivational/emotional state?
 2. Motivational State Questionnaire ([Revelle & Anderson, 1998](#)) was given participants before and after caffeine/movie/stress manipulations
 3. Data are pooled over 10 years of data (> 50 studies) in the PMC lab and available as the msqR data set
 4. Here we show how to select cases and find Cohen d ([Cohen, 1988](#))

R code

```



```

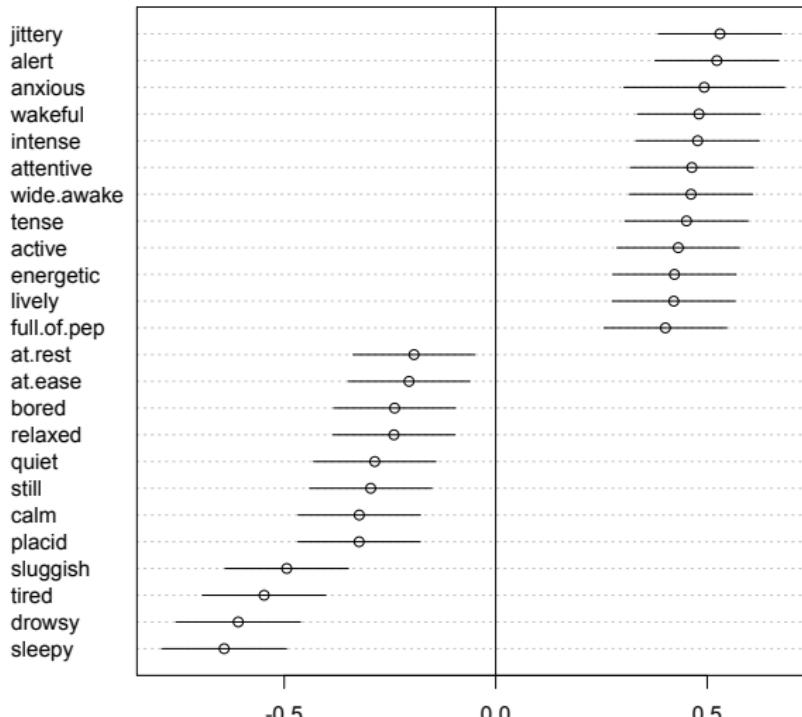
```



```

Cohen d for caffeine/placebo on msqR data

Effect of caffeine on mood



Showing sex differences in behavior

1. Athenstaedt (2003) examined Gender Role Self-Concept. She reports two independent dimensions of Male and Female behaviors.
 2. While there are large gender/sex differences on both of these dimensions, the two represent independent factors!
 3. Eagly & Revelle (2022) used these data to explore the power of aggregation when examining sex differences.
 4. Included as an example of various graphical displays.

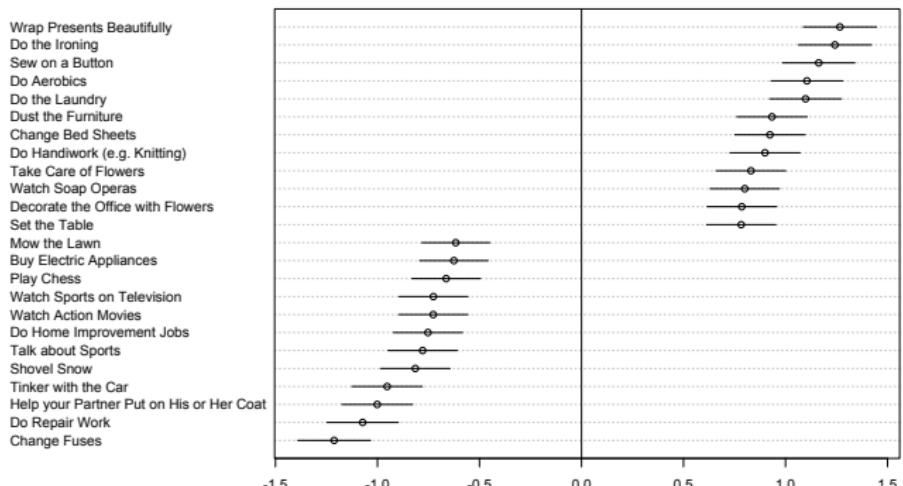
R code

```
#show error dots and ci
cd <- cohen.d(Athenstaedt[2:75], group="gender",
               dictionary=Athenstaedt.dictionary)
error.dots(cd, main="Cohen d for Athenstaedt data (with 95% CI)")
abline(v=0)

#show scatter plots and density
scatterHist(Femininity ~ Masculinity + gender, data =Athenstaedt,
cex.point=.4,smooth=FALSE, correl=FALSE,d.arrow=TRUE,
col=c("red","blue"), lwd=4, cex.main=1.5,
main="Scatter Plot and Density",cex.axis=2)
```

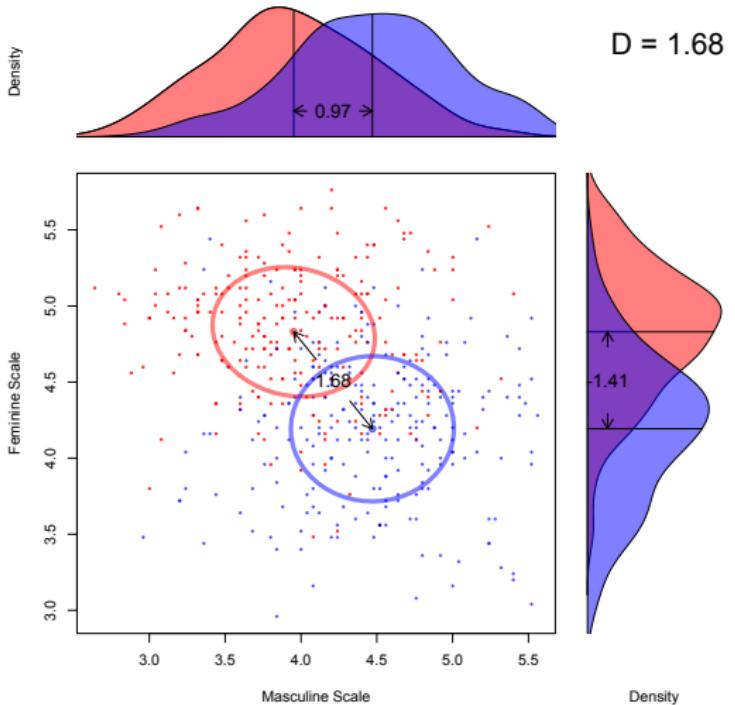
Male/female differences on GERA items Athenstaedt (2003)

Cohen d for Athenstaedt data (with 95% CI)



Male/female differences on two scales

Combined M and F scales



$r_{wg} = -.05, r = -.29$ (Athenstaedt, 2003; Eagly & Revelle, 2022)

Factor analysis, Cluster Analysis, Principal Components

1. Psychological data is typically of a high dimensionality.
 2. One solution to this problem is the factor model which interprets observed manifest observed variables in terms of unobservable, latent variables. At the data level this is of course:

$$X_i = \Lambda x_k + \Theta x_{ll} \quad (1)$$

3. At the level of covariances or correlations this is

$$\mathbf{C} \approx \Lambda\Lambda' + \Theta^2. \quad (2)$$

4. For a fixed number of factors and fixed values of Θ^2 this is solveable as a simple eigen value decomposition.
 5. However, the problem of how many factors is difficult (there is no one right answer).

Factor analysis as an iterative procedure

1. An initial estimate of communalities ($1 - \Theta^2$)
2. Find the eigen vectors (\mathbf{F}) of $\mathbf{R} - \Theta^2$
3. Find the residuals of $\mathbf{R} - \mathbf{F}'\mathbf{F}$
 - ML for maximum likelihood
 - minres for minimum residual (default)
 - pa for principal factor
 - ...
4. Set the new communalities to diagonal of $\mathbf{F}'\mathbf{F}$
5. Iterate until communalities don't change or until sum of squared residuals is a minimum or until Maximum Likelihood estimate is minimized.
6. If rotation (orthogonal) or transformation (oblique) apply the chosen algorithm.
 - "none", "varimax", "quartimax", "BentlerT", "equamax", "varimin", "geominT" and "bifactor" are orthogonal rotations.
 - "Promax", "promax", "oblimin", "simplimax", "bentlerQ", "geominQ", "biquartimin" and "cluster" are oblique transformations

How many factors – no right answer, one wrong answer

1. Statistical

- Extracting factors until the χ^2 of the residual matrix is not significant.
 - Extracting factors until the change in χ^2 from factor n to factor n+1 is not significant.

2. Rules of Thumb

- Parallel: Extracting factors until the eigenvalues of the real data are less than the corresponding eigenvalues of a random data set of the same size (*parallel analysis*) fa.parallel
 - Plotting the magnitude of the successive eigenvalues and applying the *scree test*. scree

3. Interpretability

- Extracting factors as long as they are interpretable.
 - Using the *Very Simple Structure* Criterion (VSS)
 - Using the Minimum Average Partial criterion (MAP).

4. Eigen Value of 1 rule (The worst rule)

nFactors applies many of these procedures

The number of factors problem is easy and hard

No best rule, one worst rule

“Solving the number of factors problem is easy, I do it everyday before breakfast. But knowing the right solution is harder.”
(attributed to Henry Kaiser by [Horn & Engstrom \(1979\)](#))

1. Parallel analysis (Extract factors until the eigen values are less than those of a random matrix).
 - Although a good rule for 100-500 subjects, this will not do as well with many (> 1000) subjects.
 2. [Velicer \(1976\)](#) Minimum Average Partial (MAP) is pretty good
 3. For items, the Very Simple Structure (VSS) ([Revelle & Rocklin, 1979](#)) criterion is pretty good.
 4. Multiple statistical tests, many have problems with sample size.
 - If you want few factors, run few subjects
 - If you want many factors, run many subjects
 5. One worst rule is the eigen value of 1.0 rule.

fa.parallel

R code

```
fa.parallel(bfi[1:25])  
vss(bfi[1:25])
```

```
fa.parallel(bfi[1:25])
Parallel analysis suggests that the number of factors = 6
      and the number of components = 6
```

Very Simple Structure

```
Call: vss(x = bfi[1:25])
```

VSS complexity 1 achieves a maximum of 0.58 with 4 factors

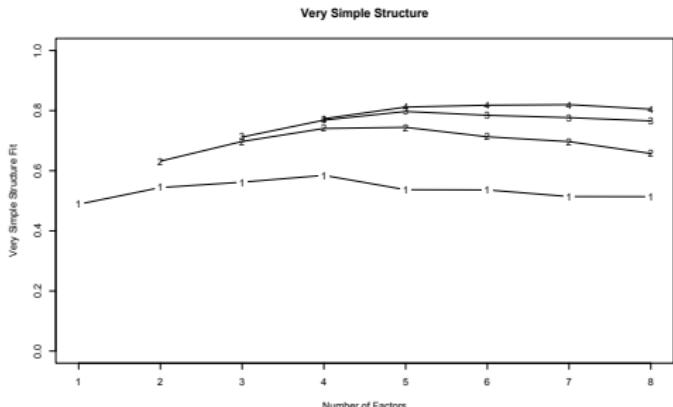
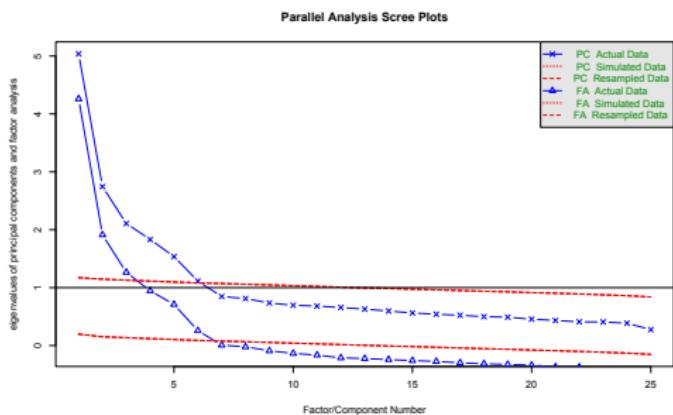
VSS complexity 2 achieves a maximum of 0.74 with 5 factors

The Velicer MAP achieves a minimum of 0.01 with 5 factors

BIC achieves a minimum of -513.09 with 8 factors

Sample Size adjusted BIC achieves a minimum of -106.39 with 8 factors

fa.parallel and vss



“The number of factors problem will break your heart”

R code

```
nFactors(bfi[1:25])
```

```
nFactors(bfi[1:25])
```

Number of factors

```
Call: vss(x = x, n = n, rotate = rotate, diagonal = diagonal, fm = fm,
n.obs = n.obs, plot = FALSE, title = title, use = use, cor = cor)
```

VSS complexity 1 achieves a maximum of 0.58 with 4 factors

VSS complexity 2 achieves a maximum of 0.74 with 5 factors

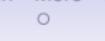
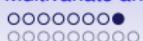
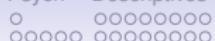
The Velicer MAP achieves a minimum of 0.01 with 5 factors.

Empirical BIC achieves a minimum of -737.9 with 8 factors.

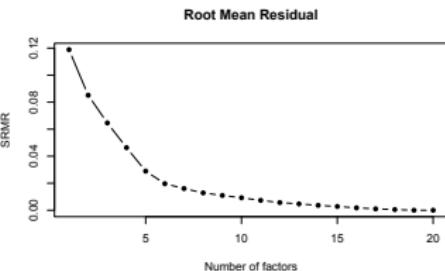
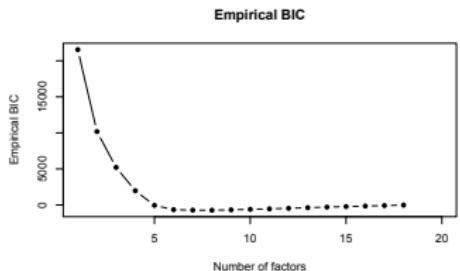
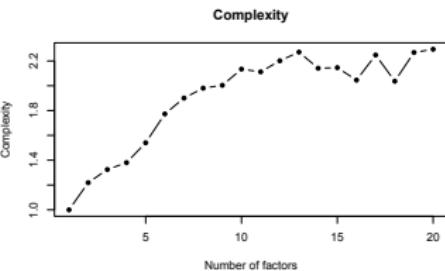
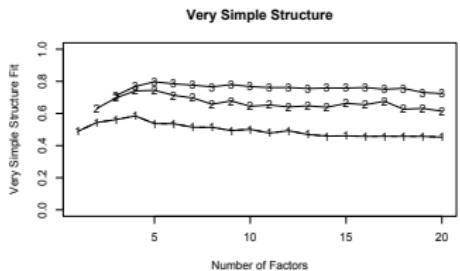
Sample Size adjusted BIC achieves a minimum of -205.18 with 12 factors

Statistics by number of factors

vss1	vss2	map	dof	chisq	prob	sqresid	fit	RMSEA	BIC	SABIC	complex	eChisq
1	0.49	0.00	0.024	275	1.2e+04	0.0e+00	25.9	0.49	0.123	9680	10554	1.0 2.4e+04
2	0.54	0.63	0.018	251	7.4e+03	0.0e+00	18.6	0.63	0.101	5370	6168	1.2 1.2e+04
3	0.56	0.70	0.017	228	5.1e+03	0.0e+00	14.6	0.71	0.087	3286	4010	1.3 7.0e+03
4	0.58	0.74	0.015	206	3.4e+03	0.0e+00	11.5	0.77	0.075	1787	2441	1.4 3.6e+03
5	0.54	0.74	0.015	185	1.8e+03	4.3e-264	9.4	0.81	0.056	341	928	1.5 1.4e+03
6	0.54	0.71	0.016	165	1.0e+03	1.8e-125	8.3	0.84	0.043	-277	247	1.8 6.5e+02
7	0.51	0.70	0.019	146	7.1e+02	1.2e-74	7.9	0.85	0.037	-451	13	1.9 4.3e+02
8	0.51	0.66	0.022	128	5.0e+02	7.1e-46	7.4	0.85	0.032	-513	-106	2.0 2.8e+02
9	0.49	0.68	0.027	111	3.8e+02	8.2e-31	7.1	0.86	0.029	-503	-150	2.0 2.0e+02
10	0.50	0.64	0.032	95	2.9e+02	4.5e-21	6.7	0.87	0.027	-468	-166	2.1 1.4e+02
11	0.48	0.65	0.039	80	1.8e+02	2.2e-09	6.4	0.87	0.021	-457	-203	2.1 9.0e+01
12	0.49	0.64	0.047	66	1.1e+02	6.9e-04	6.3	0.88	0.015	-415	-205	2.2 5.5e+01
13	0.47	0.65	0.057	53	7.6e+01	2.0e-02	6.2	0.88	0.012	-345	-176	2.3 3.7e+01
14	0.46	0.64	0.066	41	5.4e+01	7.9e-02	5.6	0.89	0.011	-271	-141	2.1 2.2e+01



How many factors? What ever you want



fa (from the help page)

Exploratory Factor analysis using MinRes (minimum residual) as well as EFA by Principal Axis, Weighted Least Squares or Maximum Likelihood

Among the many ways to do latent variable exploratory factor analysis (EFA), one of the better is to use Ordinary Least Squares (OLS) to find the minimum residual (minres) solution. This produces solutions very similar to maximum likelihood even for badly behaved matrices. A variation on minres is to do weighted least squares (WLS). Perhaps the most conventional technique is principal axes (PAF). An eigen value decomposition of a correlation matrix is done and then the communalities for each variable are estimated by the first n factors. These communalities are entered onto the diagonal and the procedure is repeated until the sum(diag(r)) does not vary. Yet another estimate procedure is maximum likelihood. For well behaved matrices, maximum likelihood factor analysis (either in the fa or in the factanal function) is probably preferred. Bootstrapped confidence intervals of the loadings and interfactor correlations are found by fa with n.iter > 1.

More important output

R code

```
f5
diagram(f5, main="5 factors of the bfi")
plot(f5) #an alternative way to show the results
biplot(f5 ) #show a biplot
```

Mean item complexity = 1.5

Test of the hypothesis that 5 factors are sufficient.

df null model = 300 with the objective function = 7.23 with Chi Square = 20163.79
df of the model are 185 and the objective function was 0.65

The root mean square of the residuals (RMSR) is 0.03

The df corrected root mean square of the residuals is 0.04

The harmonic n.obs is 2762 with the empirical chi square 1392.16 with prob < 5.6e-184
The total n.obs was 2800 with Likelihood Chi Square = 1808.94 with prob < 4.3e-264

Tucker Lewis Index of factoring reliability = 0.867

RMSEA index = 0.056 and the 90% confidence intervals are 0.054 0.058

BIC = 340.53

Fit based upon off diagonal values = 0.98

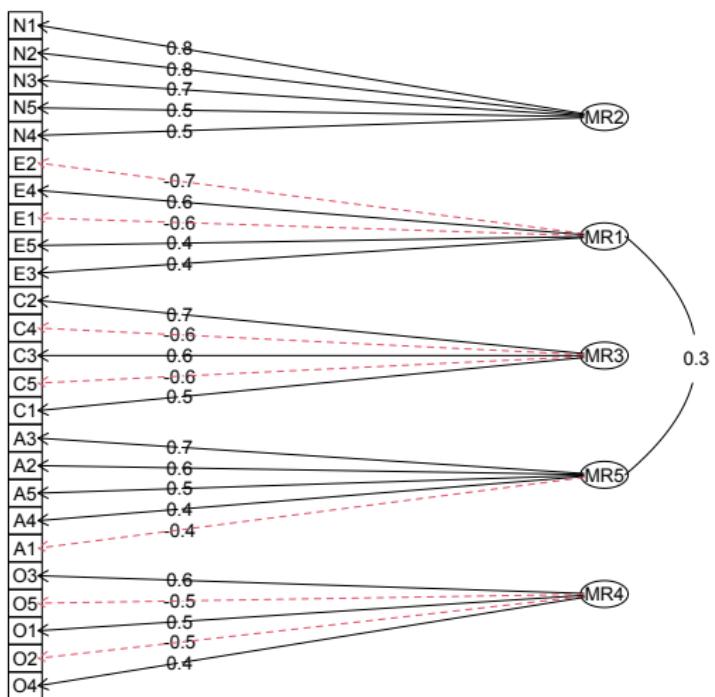
Measures of factor score adequacy

	MR2	MR1	MR3	MR5	MR4
Correlation of (regression) scores with factors	0.92	0.89	0.88	0.88	0.84
Multiple R square of scores with factors	0.85	0.79	0.77	0.77	0.71
Minimum correlation of possible factor scores	0.70	0.59	0.54	0.54	0.42
>					



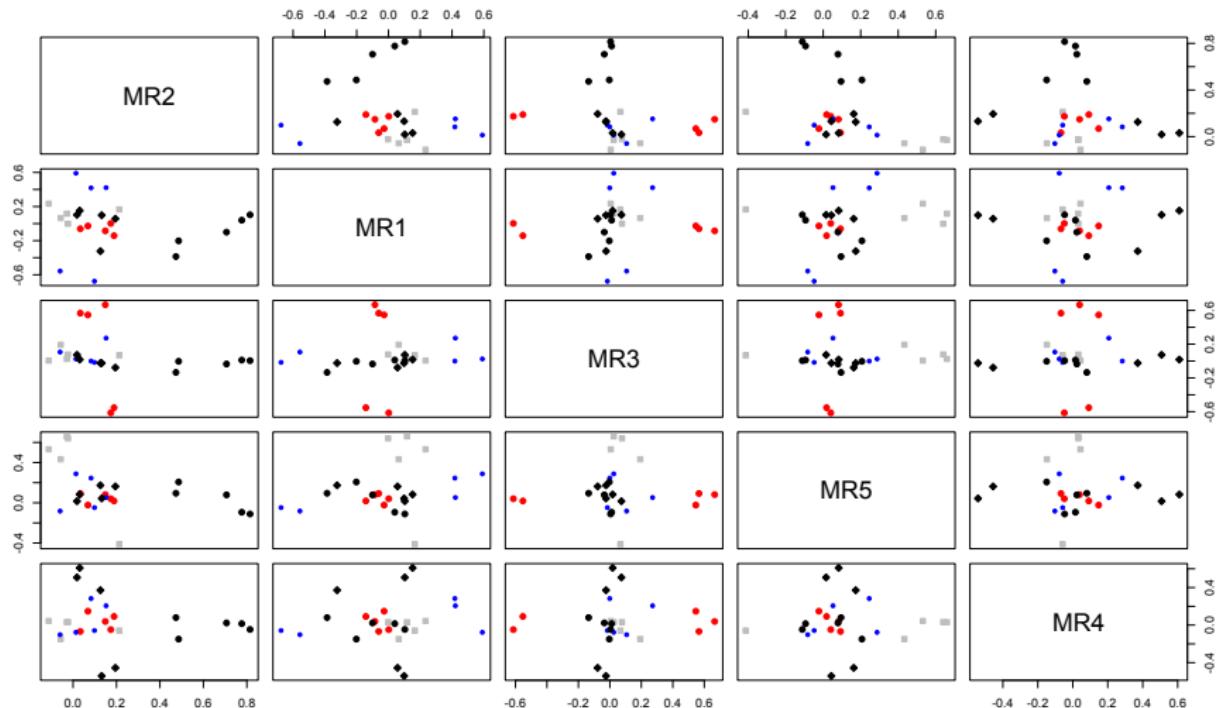
Use the diagram to show the structure

5 factors of the bfi

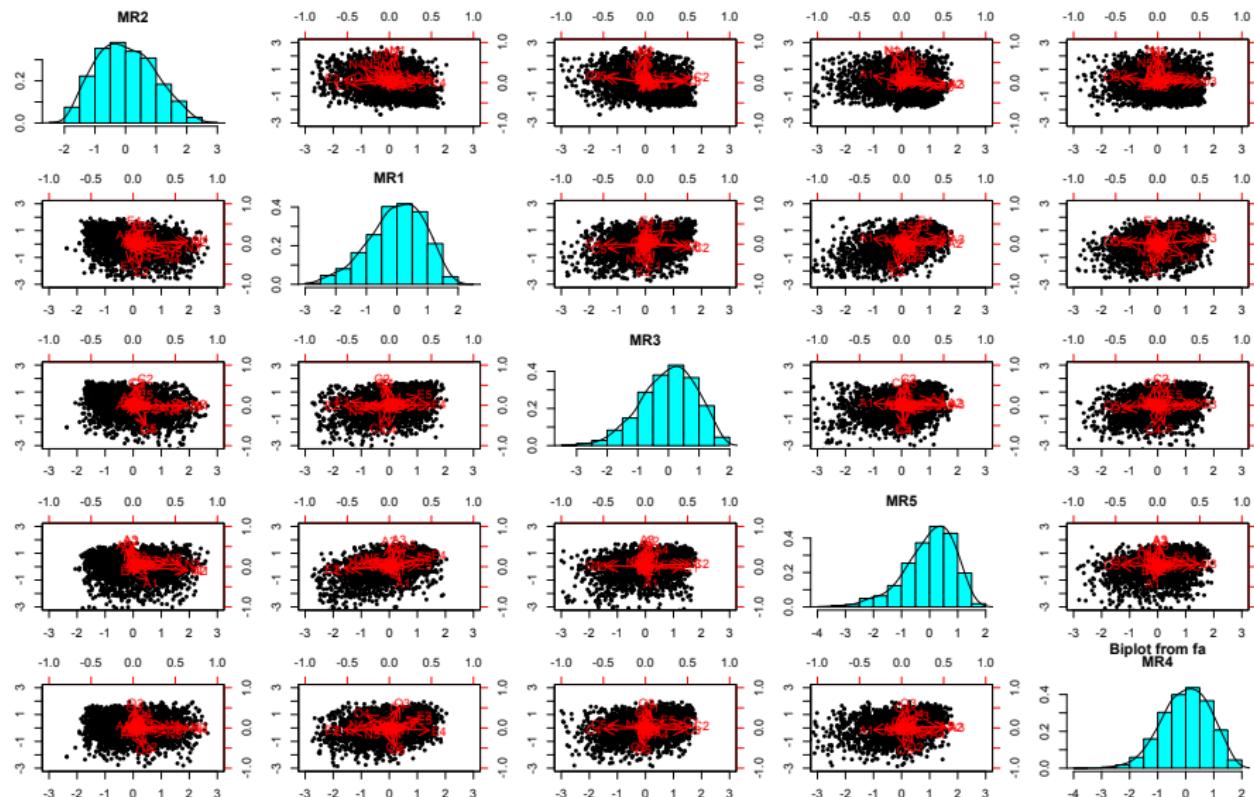


Use the plot to show the loadings in a different fashion

Factor Analysis



Use the biplot to show the loadings in a different fashion



Cluster analysis as an alternative to factor analysis

The ICLUS algorithm was developed for the construction of internally consistent scales from items [Revelle \(1979\)](#)

1. Form the proximity (correlation) matrix
 2. Find the most similar pair of items
 3. Combine them into a new scale,
 4. Recalculate the correlation matrix
 5. Repeat steps 2-4 until various criteria are met

alpha Coefficient α of the composite fails to increase (rarely happens)

beta Coefficient *beta* (the worst split half reliability) fails to increase

iclust Is meant for forming scales from items and is a useful guide to the structure of a test.

iclust **Revelle (1979)**

R code

```
ic <- iclust(bfi[1:25])  
summary(ic)
```

```
ICLUST (Item Cluster Analysis) Call: iclust(r.mat = bfi[1:25])  
ICLUST
```

Purified Alpha:
C20 C16 C15 C21
0.80 0.81 0.73 0.61

Guttman Lambda6*

Original Beta:

Cluster size:
C20 C16 C15 C21
10 5 5 5

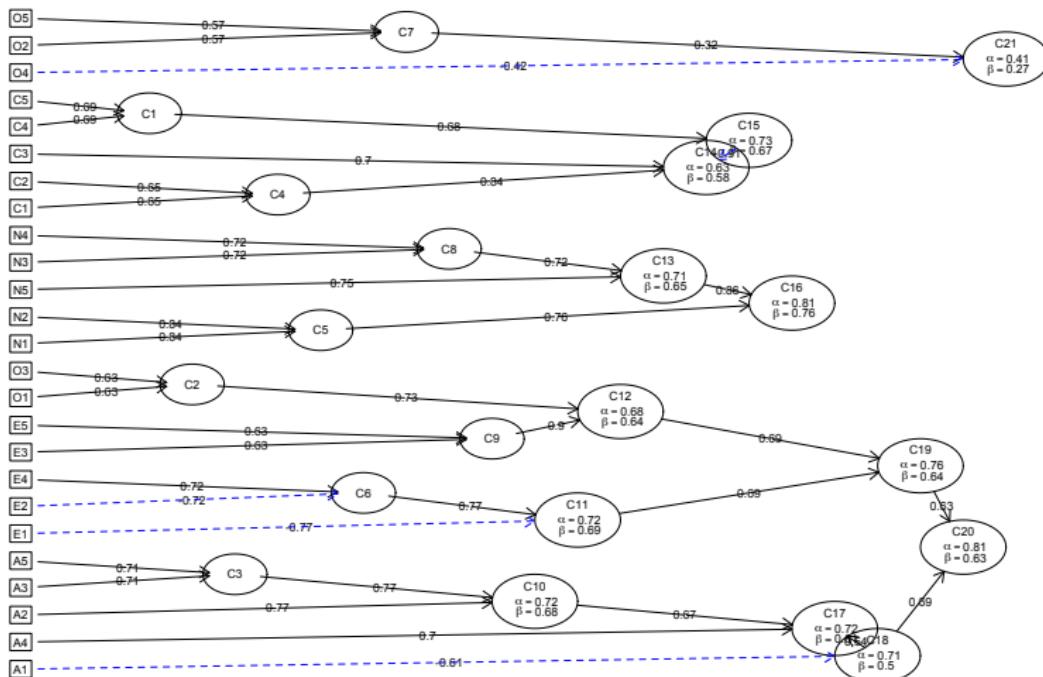
```

Purified scale intercorrelations
reliabilities on diagonal
correlations corrected for attenuation above diagonal:
      C20    C16    C15    C21
C20  0.80 -0.291 -0.40 -0.33
C16 -0.24  0.815  0.29  0.11
C15 -0.30  0.221  0.73  0.30
C21 -0.23  0.074  0.20  0.61

```

Hierarchical cluster analysis of items using iclust

ICLUST



Multiple levels of factors

1. Hierarchical/higher order models (Jensen & Weng, 1994)
 - Simulated data to match Jensen & Weng (1994)
 - Ability data from the ICAR Condon & Revelle (2014)
 - Size data from the United States Airforce
 - Hierarchical solution of the SAPA Personality Inventory (spi)
(Condon, 2018)
 2. Bifactor models (Holzinger & Swineford, 1937; Reise, 2012; Rodriguez, Reise & Haviland, 2016)
 - Schmid & Leiman (1957) introduced a transformation of a higher order model into a bifactor model.
 - Constraints on the factor loadings given the structure.
 3. Comparing factors at different levels of n factors using the bassAckward algorithm (Goldberg, 2006; Waller, 2007)
 - Two levels of factors from the SAPA Personality Inventory (spi)
(Condon, 2018)
 4. The SAPA Personality Inventory (spi) (Condon, 2018) data set has 135 items plus 10 criteria variables for 4,000 participants.
 - It was developed from 696 IPIP items to represent 200 broad and narrow public domain measures of personality.

Simulating 9 variables from Jensen & Weng (1994)

R code

```
jensen <- sim.hierarchical() #the default values are Jensen-Weng
f3 <- fa(jensen, 3)
om<- omega(jensen)
diagram(om, sl=FALSE); diagram(om) #default is to do Schmid-Leiman
```

```

Factor Analysis using method = minres
Call: fa(r = jensen, nfactors = 3)
Standardized loadings (pattern matrix) based upon correlation matrix
    MR1 MR3 MR2   h2   u2 com
V1 0.8 0.0 0.0 0.64 0.36   1
V2 0.7 0.0 0.0 0.49 0.51   1
V3 0.6 0.0 0.0 0.36 0.64   1
V4 0.0 0.7 0.0 0.49 0.51   1
V5 0.0 0.6 0.0 0.36 0.64   1
V6 0.0 0.5 0.0 0.25 0.75   1
V7 0.0 0.0 0.6 0.36 0.64   1
V8 0.0 0.0 0.5 0.25 0.75   1
V9 0.0 0.0 0.4 0.16 0.84   1

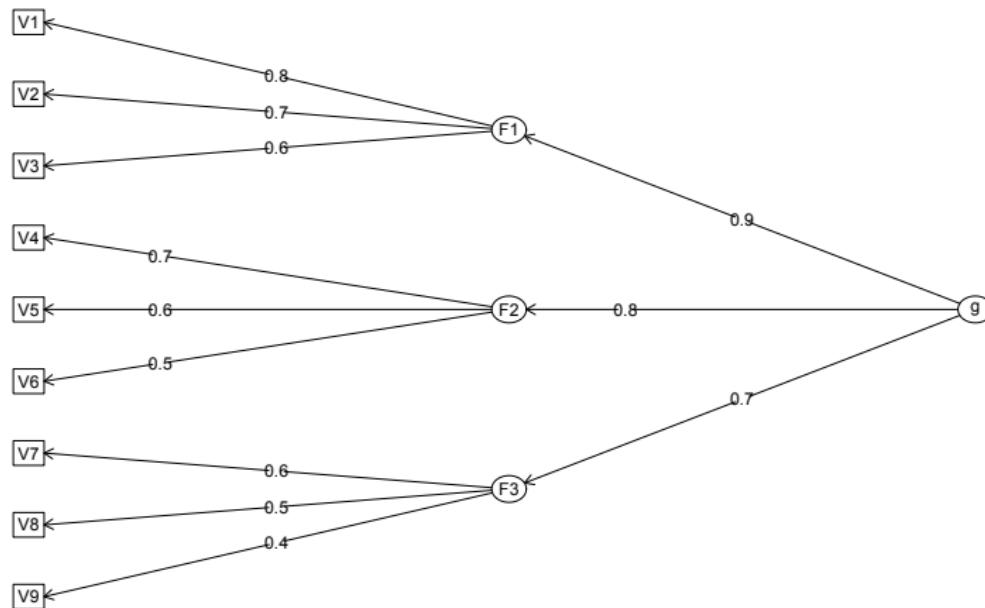
```

	MR1	MR3	MR2
SS loadings	1.49	1.10	0.77
Proportion Var	0.17	0.12	0.09
Cumulative Var	0.17	0.29	0.37
Proportion Explained	0.44	0.33	0.23
Cumulative Proportion	0.44	0.77	1.00

With factor correlations of
 MR1 MR3 MR2
 MR1 1.00 0.72 0.63
 MR3 0.72 1.00 0.56
 MR2 0.63 0.56 1.00

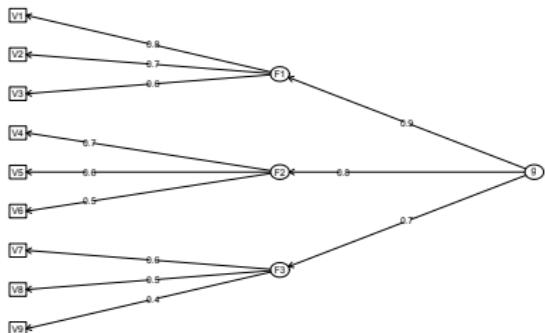
A higher order factor representation

Hierarchical (multilevel) Structure

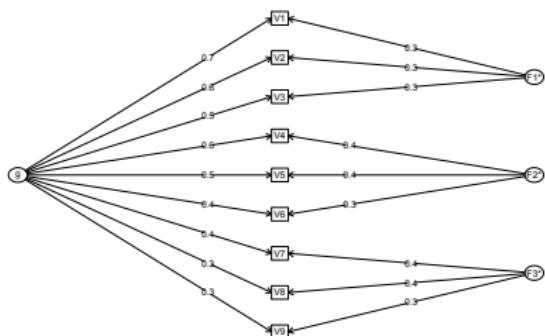


Schmid & Leiman (1957) transformation to a bifactor model

Hierarchical (multilevel) Structure



Omega with Schmid Leiman Transformation



Unfortunately, the bifactor rotation does not capture the right structure

R code

```
f4 <- fa(jensen.4, rotate="bifactor")
```

```

Factor Analysis using method = minres
Call: fa(r = jensen, nfactors = 4, rotate = "bifactor")
Standardized loadings (pattern matrix) based upon correlation matrix
    MR1    MR3    MR2    MR4    h2   u2 com
V1  0.79 -0.03 -0.09  0.02  0.63  0.37 1.0
V2  0.70 -0.05 -0.09 -0.06  0.51  0.49 1.1
V3  0.60 -0.03 -0.07  0.12  0.38  0.62 1.1
V4  0.53  0.45  0.01  0.00  0.49  0.51 2.0
V5  0.46  0.39  0.01  0.00  0.36  0.64 2.0
V6  0.38  0.32  0.00  0.00  0.25  0.75 2.0
V7  0.43  0.01  0.42  0.00  0.36  0.64 2.0
V8  0.36  0.01  0.35  0.00  0.25  0.75 2.0
V9  0.29  0.01  0.28  0.00  0.16  0.84 2.0

    MR1    MR3    MR2    MR4
SS loadings  2.50  0.47  0.39  0.02
Proportion Var 0.28  0.05  0.04  0.00
Cumulative Var 0.28  0.33  0.37  0.38
Proportion Explained 0.74  0.14  0.12  0.01
Cumulative Proportion 0.74  0.88  0.99 1.00

```

Another case: the ICAR 16

R code

```
om.icar <- omega(icar, 4)
```

Schmid Leiman Factor loadings greater than 0.2

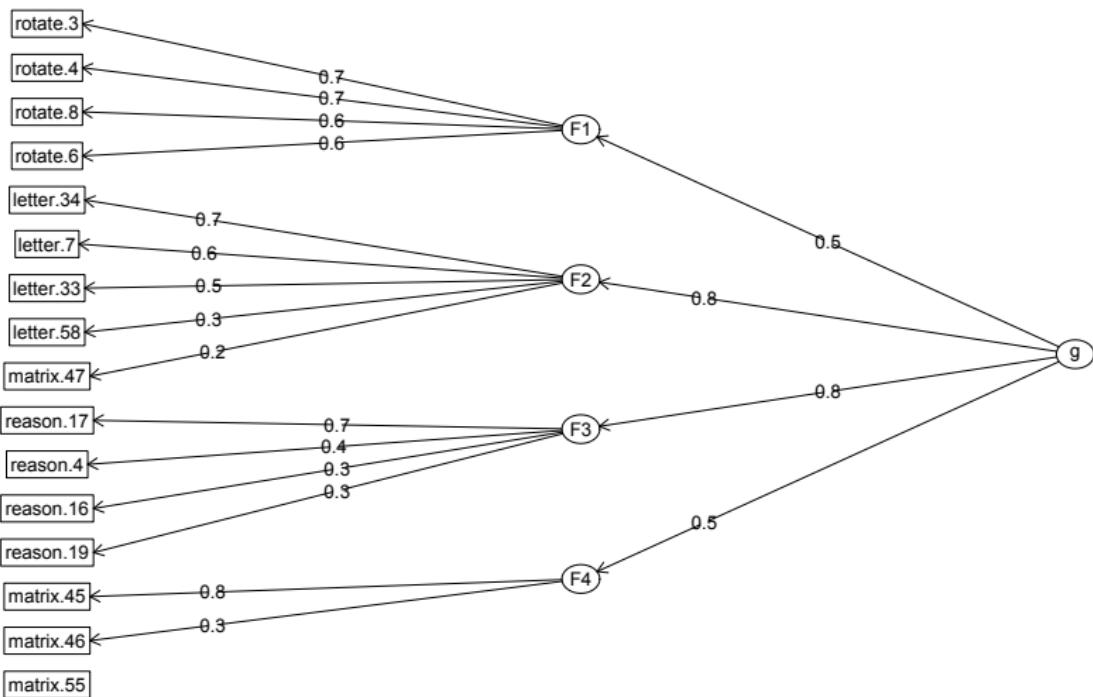
	g	F1*	F2*	F3*	F4*	h2	u2	p2
reason.4	0.50			0.28		0.35	0.65	0.74
reason.16	0.42			0.21		0.23	0.77	0.76
reason.17	0.55			0.46		0.51	0.49	0.59
reason.19	0.44			0.21		0.25	0.75	0.78
letter.7	0.51		0.35			0.39	0.61	0.68
letter.33	0.46		0.31			0.31	0.69	0.69
letter.34	0.53		0.39			0.43	0.57	0.65
letter.58	0.47		0.20			0.28	0.72	0.78
matrix.45	0.40				0.64	0.57	0.43	0.28
matrix.46	0.40				0.26	0.24	0.76	0.65
matrix.47	0.43					0.23	0.77	0.79
matrix.55	0.29					0.13	0.87	0.66
rotate.3	0.36	0.60				0.50	0.50	0.26
rotate.4	0.41	0.60				0.53	0.47	0.32
rotate.6	0.40	0.49				0.41	0.59	0.39
rotate.8	0.33	0.54				0.41	0.59	0.27

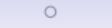
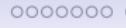
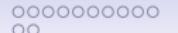
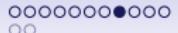
With Sums of squares of:

g	F1*	F2*	F3*	F4*
3.05	1.31	0.47	0.40	0.53

omega of the ability items

Hierarchical (multilevel) Structure



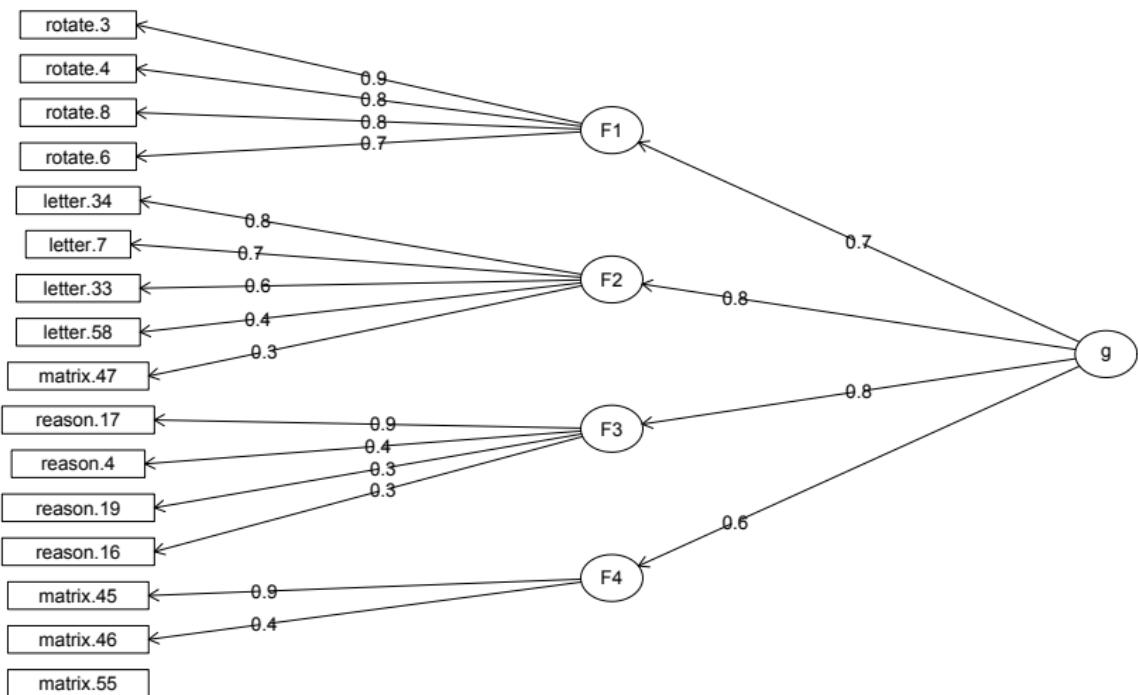


Pearson, polychoric and tetrachoric correlations

1. As we know, Pearson correlations are appropriate for continuous data.
2. But with categorical or dichotomous data, the correlations are attenuated.
3. The tetrachoric correlation estimates what the Pearson would be with continuous data.
 - Tetrachoric and polychoric correlations are thus estimates of the *latent* correlation assuming bivariate normality.
 - Appropriate for determining the structure of correlations, inappropriate for estimates of reliability.
4. The fa, omega functions have an option to find the tetrachoric/polychoric correlations before factoring.
5. Particularly appropriate for dichotomous variables (e.g. the ICAR example, ability)

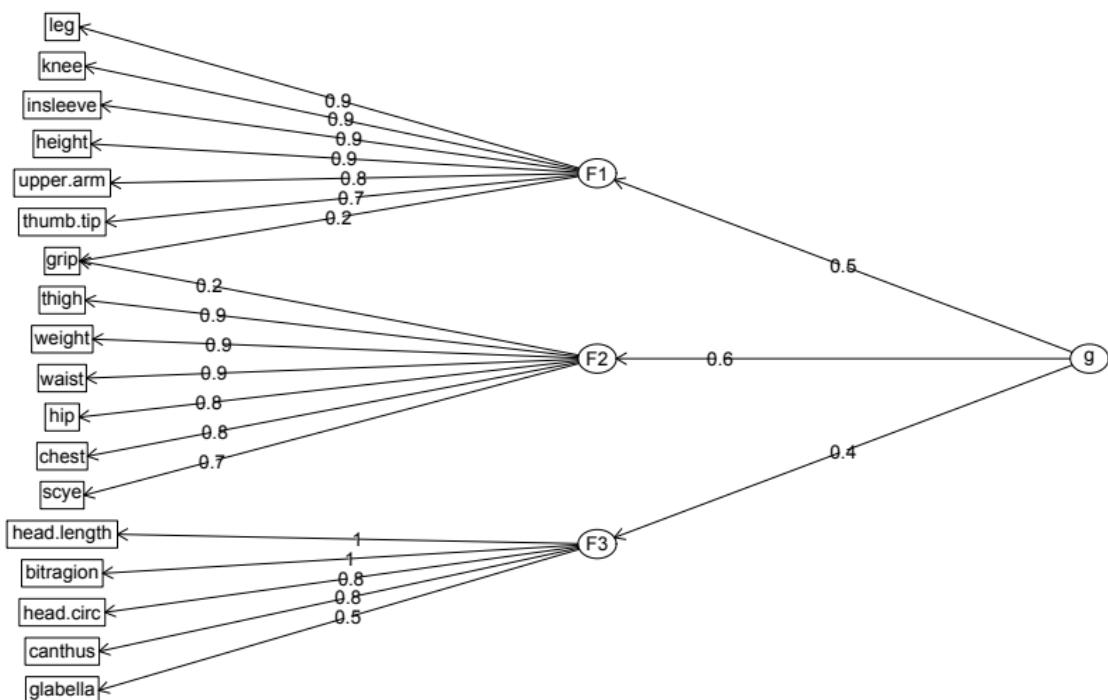
omega of the ability items using tetrachoric correlations

Hierarchical/multilevel Structure using tetrachoric correlations



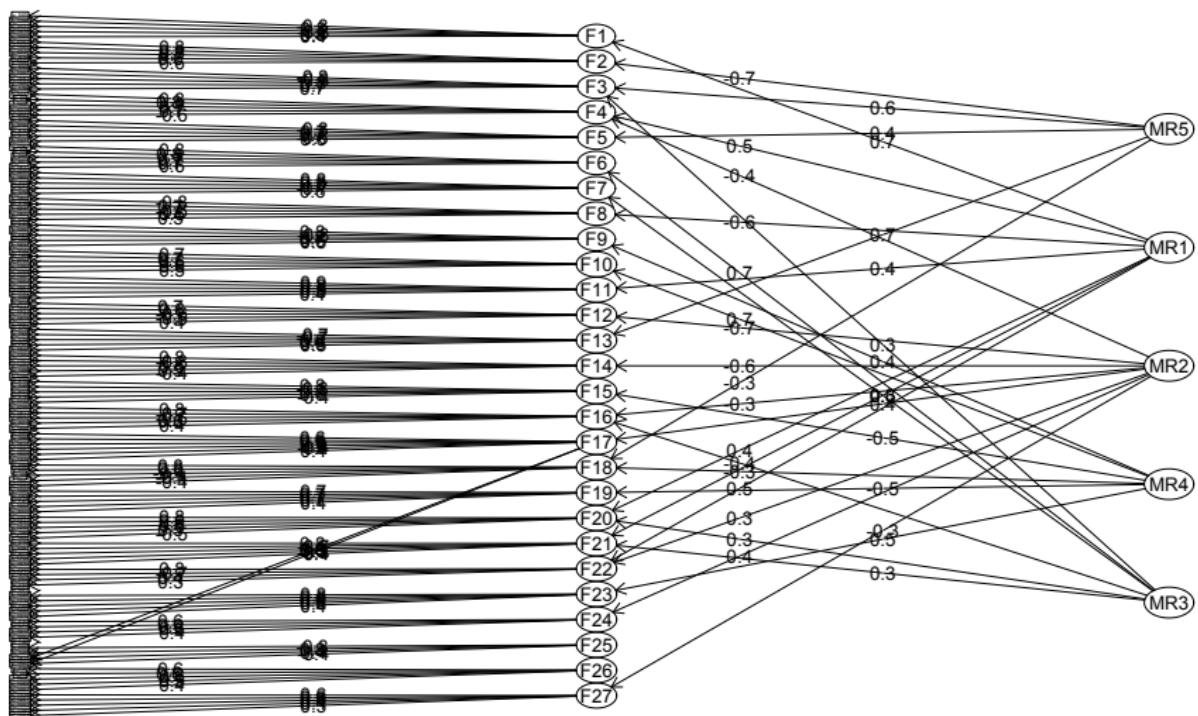
Is there a general factor of body size? The USAF data set

g of bodysize?



fa.hierarchical solution of the spi of the spi items

Hierarchical (multilevel) Structure





●

○

○

○

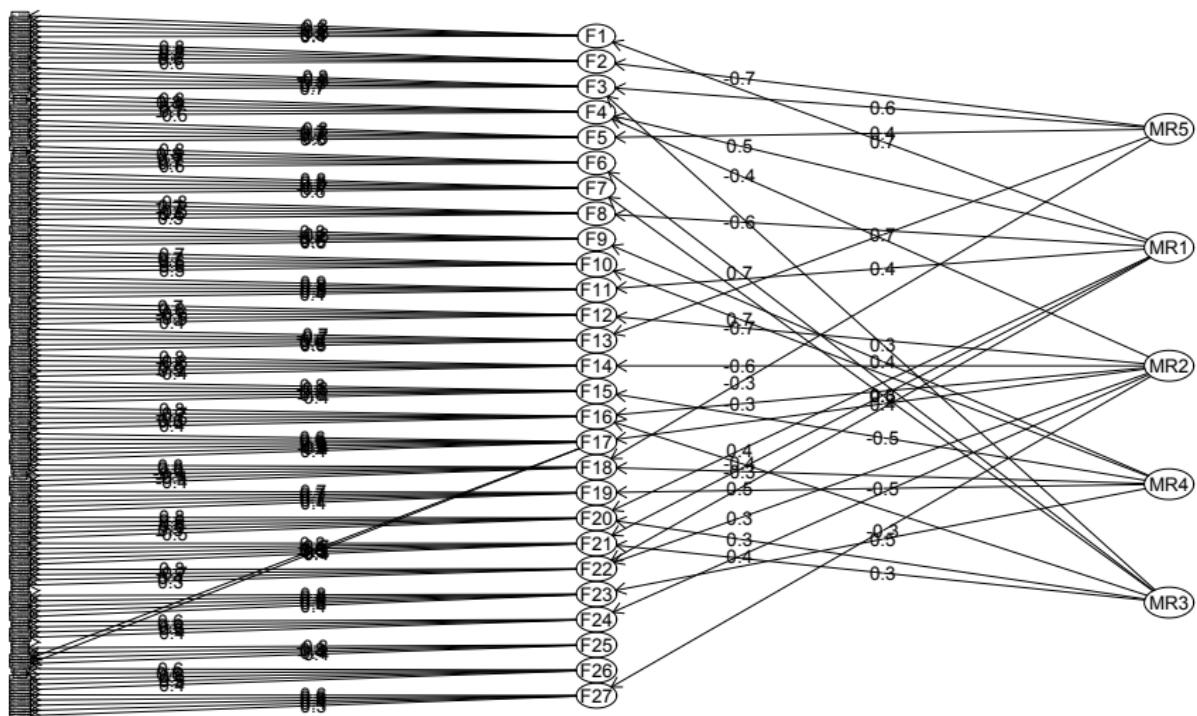
The “Bass-Ackwards” algorithm

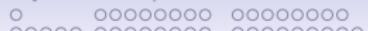
1. Goldberg (2006) described a hierarchical factor structure organization from the “top down”.
 - The original idea was to do successive factor analyses from 1 to nf factors organized by factor score correlations from one level to the next.
2. Waller (2007) discussed a simple way of doing this for components without finding the scores.
3. Using the factor correlations (from Gorsuch, 1983) to organize factors hierarchically results may be organized at many different levels.
4. The algorithm may be applied to principal components (pca) or to true factor analysis.
5. Implemented as bassAckward.
6. The solutions should not be confused with a hierarchical solution where the higher order factors are factors of the lower order factors.



bassAckward solution of the spi items for 5 and 27 factors

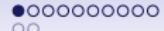
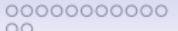
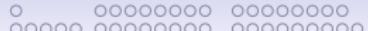
Hierarchical (multilevel) Structure





Scale Construction

1. *psych* was specifically designed for the problem of reading and describing sets of items and then forming unit weighted scales from these items.
2. The advantage of scales formed from unit weighted items rather than factor weights is that they are more robust to sample variation ([Widaman & Revelle, 2022](#)).
3. Although there are functions to combine a set of items into just one scale alpha the more typical problem is form multiple scales e.g., `scoreItems`.
4. `fastScore` will scale scores without any accompanying statistics, but the more typical case is to use `.pfunscoreItems`.
5. To find scales based upon Item Response Theory, use `scoreIrt`



Example data sets

1. The sai represents 3,032 participants on 20 state anxiety items with 1229 participants who took it twice, 1047 with three measures, and 70 with four measures.
2. The epi represents 3570 participants for the 57 items of the Eysenck Personality Inventory from the early 1990s at the PMC lab.
3. An additional data set (epiR) has test and retest information for 474 participants.
4. The Motivational State Questionnaire msqR ([Revelle & Anderson, 1998](#)) contains 75 mood items for 3032 unique participants . 2753 took it at least twice, 446 three times, and 181 four times.

Scoring scales

1. For one set of items for one scale use alpha
 - Will warn if item x total correlations are negative and encourage
 - Using the check.keys option to reverse score negatively keyed items
 2. More typical is to specify a keys.list of multiple keys each with multiple items.
 - Negatively keyed items are reversed scored by subtracting from the maximum possible item score - minimum possible item score
 - Scale scores are expressed as the *mean* item response, although *sum* scores is also an option,
 - Missing items scores can be *imputed* by means, medians, or ignored.
 3. Most scoring functions return scores as well as statistics for the scales.
 4. scoreFast and scoreVeryFast just return the scores.

Example keys list

R code

```
sai.keys <- list(sai = c("tense", "regretful", "upset", "worrying", "anxious", "nervous",
"jittery", "high.strung", "worried", "rattled", "-calm",
"-secure", "-at.ease", "-rested", "-comfortable", "-confident", "-relaxed", "-content",
"-joyful", "-pleasant"),
sai.p = c("calm", "at.ease", "rested", "comfortable", "confident", "secure", "relaxed",
"content", "joyful", "pleasant"),
sai.n = c( "tense", "anxious", "nervous", "jittery", "rattled", "high.strung",
"upset", "worrying", "worried", "regretful")
)
sai.keys
```

```
$sai
[1] "tense"      "regretful"    "upset"       "worrying"     "anxious"      "nervous"
[7] "jittery"     "high.strung"  "worried"     "rattled"     "-calm"       "-secure"
[13] "-at.ease"    "-rested"     "-comfortable" "-confident"   "-relaxed"    "-content"
[19] "-joyful"     "-pleasant"
```

```
$sai.p
[1] "calm"        "at.ease"      "rested"      "comfortable" "confident"   "secure"
"relaxed"      "content"     "joyful"     "pleasant"
```

```
$sai.n
[1] "tense"      "anxious"     "nervous"    "jittery"     "rattled"     "high.strung"
"upset"        "worrying"    "worried"    "regretful"
```

Some keys.list are part of the data set

R code

epi.keys

```

epi.keys
$E
[1] "V1"    "V3"    "V8"    "V10"   "V13"   "V17"   "V22"   "V25"   "V27"   "V39"
[11] "V44"   "V46"   "V49"   "V53"   "V56"   "-V5"   "-V15"  "-V20"  "-V29"  "-V32"
[21] "-V34"  "-V37"  "-V41"  "-V51"

$N
[1] "V2"    "V4"    "V7"    "V9"    "V11"   "V14"   "V16"   "V19"   "V21"   "V23"   "V26"   "V28"
[13] "V31"   "V33"   "V35"   "V38"   "V40"   "V43"   "V45"   "V47"   "V50"   "V52"   "V55"   "V57"

$L
[1] "V6"    "V24"   "V36"   "-V12"  "-V18"  "-V30"  "-V42"  "-V48"  "-V54"

$Imp
[1] "V1"    "V3"    "V8"    "V10"   "V13"   "V22"   "V39"   "-V5"   "-V41"

$Soc
[1] "V17"   "V25"   "V27"   "V44"   "V46"   "V53"   "-V11"  "-V15"  "-V20"  "-V29"
[11] "-V32"  "-V37"  "-V51"

```

Dictionaries

1. Referring to item numbers is not convenient for discussing results.
 2. Thus, it is possible to create a dictionary of the items.
 3. A dictionary can be prepared outside of R by forming a spreadsheet including at least one column labeled “content” and with rownames for the item number or name. Other columns can specify the item source, or anything interesting.

```
headTail(epi.dictionary)
```

	Content
V1	Do you often long for excitement?
V2	Do you often need understanding friends to cheer you up?
V3	Are you usually carefree?
V4	Do you find it very hard to take no for an answer?
...	<NA>
V54	Do you sometimes talk about things you know nothing about?
V55	Do you worry about your health?
V56	Do you like playing pranks on others?
V57	Do you suffer from sleeplessness?

Using a keys list and a dictionary to show content

R code

```
lookupFromKeys(epi.keys, epi.dictionary, n=2)
```

SE

Content

V1 Do you often long for excitement?
V3 Are you usually carefree?

SN

Content

V2 Do you often need understanding friends to cheer you up?
V4 Do you find it very hard to take no for an answer?

SI

V6 If you say you will do something do you always keep your promise,
no matter how inconvenient it might be to do so?
V24 Are all your habits good and desirable ones?

\$tmp

Content

V1 Do you often long for excitement?
V3 Are you usually carefree?

550c

Content

V17 Do you like going out a lot?

V25 Can you usually let yourself go and enjoy yourself a lot at a lively party?

Or show the items for just one scale (as a way of checking the keys)

R code

```
lookupFromKeys(epi.keys, epi.dictionary) $Imp
```

```
lookupFromKeys(epi.keys, epi.dictionary) $Imp
```

Content

Using scoreItems on the epi dataset

R code

```
scales <- scoreItems(epi.keys, epi)
overlap <- scoreOverlap(epi.keys, epi)
```

Scale intercorrelations corrected for attenuation
raw correlations below the diagonal, (unstandardized) alpha on the diagonal
corrected correlations above the diagonal:

	E	N	L	Imp	Soc
E	0.73	-0.228	-0.40	1.211	1.19
N	0.17	-0.762	0.39	0.005	0.32

Imp 0.71 0.015 -0.16 0.478 0.56 <- note that the imp and Soc scales
Soc 0.86 -0.250 -0.18 0.330 0.73 <- overlapping items with the E scale

```
> overlap <- scoreOverlap(epi.keys, epi)
```

7

```
> summary(overlap)
```

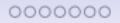
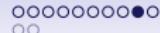
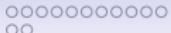
```
Call: scoreOverlap(keys = epi.keys, r = epi)
```

Scale intercorrelations adjusted for item overlap

Scale intercorrelations corrected for attenuation

raw correlations (corrected for overlap) below the diagonal, (standardized) alpha on the diagonal, and corrected (for overlap and reliability) correlations above the diagonal:

	E	N	L	Imp	Soc
E	0.73	-0.23	-0.38	0.799	0.94
N	-0.18	0.80	-0.28	0.049	-0.31
L	-0.22	-0.17	0.45	-0.311	-0.30
Imp	0.47	0.03	-0.14	0.474	0.54
Soc	0.68	-0.24	-0.17	0.320	0.73



But what if we have overlapping scales?

1. Sometimes we are interested in how higher order scales relate to lower order scales.
2. The problem is, the items overlap.
3. Some people solve this problem by dropping the overlapping items. But this changes the meaning of the scales.
4. A fairly straight forward procedure is estimate the overlapping variances with the best estimate of shared (common) variance, similar to what is done when finding coefficient α .
5. Need to do this on the correlation matrix of the items, not the raw data.
6. See ?scoreOverlap

A small part of the output for scoreItems

R code

names (scales)

```
names(scales)
[1] "scores"           "missing"          "alpha"            "av.r"
[5] "sn"               "n.items"          "item.cor"        "cor"
[9] "corrected"        "G6"                "item.corrected" "response.freq"
[13] "raw"              "ase"               "med.r"           "keys"
[17] "MIMS"             "MIMT"              "Call"

dim(scales$scores)
[1] 3570      5
```

By default, scoreItems imputes item medians for missing data

R code

```
describe(scales$scores)
scales <- scoreItems(epi.keys, epi, impute="none")
describe(scales$scores)
```

```

describe(scales$scores)
  vars   n mean   sd median trimmed  mad min max range skew kurtosis se
E     1 3570 1.46 0.17    1.46    1.46 0.19 1.04    2  0.96  0.26 -0.28  0
N     2 3570 1.54 0.19    1.54    1.55 0.19 1.00    2  1.00 -0.06 -0.42  0
L     3 3570 1.73 0.18    1.78    1.74 0.16 1.00    2  1.00 -0.57 -0.14  0
Imp   4 3570 1.51 0.20    1.56    1.51 0.16 1.00    2  1.00 -0.10 -0.57  0
Soc   5 3570 1.45 0.21    1.38    1.44 0.23 1.00    2  1.00  0.37 -0.46  0
> scales <- scoreItems(epi.keys, epi, impute="none")
> describe(scales$scores)
  vars   n mean   sd median trimmed  mad min max range skew kurtosis se
E     1 3516 1.46 0.17    1.46    1.46 0.19 1    2    1  0.20 -0.34  0
N     2 3514 1.54 0.19    1.54    1.55 0.19 1    2    1 -0.06 -0.47  0
L     3 3510 1.73 0.18    1.78    1.74 0.16 1    2    1 -0.56 -0.10  0
Imp   4 3516 1.52 0.20    1.56    1.52 0.16 1    2    1 -0.16 -0.51  0
Soc   5 3509 1.45 0.22    1.46    1.44 0.23 1    2    1  0.34 -0.52  0

```

The structure of scales can be found from correlation matrices

1. The typical use of scoring scales is from raw data.
 2. But for those of us interested in large data matrices with lots of missing data, it is convenient to score from the correlation matrix level.
 3. If we just care about the correlations of composite scales, the correlations are adequate.
 4. Given a $N \times n$ data matrix of deviation scores, for N subjects on n items, $_N\mathbf{X}_n$, the covariance matrix of the n items, $_n\mathbf{C}_n$, is just

$$n\mathbf{C}_n = \mathbf{X}\mathbf{X}' * (N - 1)^{-1},$$

with variances, σ_i^2 , on the diagonal of \mathbf{C} and item by item covariances, σ_{ij} , off the diagonal.

5. The covariances of scales are just

$$_k \mathbf{C} \mathbf{s}_k = {}_k \mathbf{K}'_{n,n} \mathbf{C}_{n,n} \mathbf{K}_k; \quad (3)$$

Scales from correlations

R code

```
R <- lowerCor(epi, show=FALSE)
fromCors <- scoreItems(epi.keys, R)
summary(fromCors)
```

```
Call: scoreOverlap(keys = epi.keys, r = R)
```

Scale intercorrelations adjusted for item overlap

Scale intercorrelations corrected for attenuation

raw correlations (corrected for overlap) below the diagonal, (standardized) alpha on the diagonal, and corrected (for overlap and reliability) correlations above the diagonal:

	E	N	L	Imp	Soc
E	0.73	-0.23	-0.38	0.799	0.94
N	-0.18	0.80	-0.28	0.049	-0.31
L	-0.22	-0.17	0.45	-0.311	-0.30
Imp	0.47	0.03	-0.14	0.474	0.54
Soc	0.68	-0.24	-0.17	0.320	0.73

Multiple types of reliability

1. Internal consistency estimates
 - α , λ_6 , use the alpha or scoreItems functions
 - $\omega_{hierarchical}$ and ω_{total} use the omega function
 2. IntraClass coefficients
 - ICC
 3. Rater agreement use kappa function
 4. Test Retest reliability

For the next examples we will use a built in data set

1. bfi consists of 25 personality items measuring 5 factors as well as some demographics.
 2. The data were collected as part of the SAPA project and have 2,800 subjects.
 3. For help on this data set, ?bfi
 4. To see all of the *psych* data sets: data(package="psych")

First, we intentionally misspecify the data [R code]

R code

```
alpha(bfi[1:5]) #score the first five items
```

Some items (A1) were negatively correlated with the total scale and probably should be reversed.

To do this, run the function again with the 'check.keys=TRUE' option.

Reliability analysis

```
Call: alpha(x = bfi[1:5])
```

raw_alpha	std.alpha	G6(smc)	average_r	S/N	ase	mean	sd
0.43	0.46	0.53	0.15	0.85	0.016	4.2	0.74

lower alpha upper 95% confidence boundaries
 0.4 0.43 0.46

Reliability if an item is dropped:

	raw_alpha	std.alpha	G6(smc)	average_r	S/N	alpha se
A1	0.72	0.73	0.67	0.398	2.64	0.0087
A2	0.28	0.30	0.39	0.097	0.43	0.0219
A3	0.18	0.21	0.31	0.061	0.26	0.0249
A4	0.25	0.31	0.44	0.099	0.44	0.0229
A5	0.21	0.24	0.36	0.072	0.31	0.0238

Item statistics

n raw.r std.r r.cor r.drop mean sd

Try it again. Turn on automatic reversals. Get the scores

R code

```
scores <- alpha(bfi[1:5], check.keys =TRUE)
```

```
alpha(bfi[1:5], check.keys =TRUE)
```

Reliability analysis

```
Call: alpha(x = bfi[1:5], check.keys = TRUE)
```

raw_alpha	std.alpha	G6(smc)	average_r	S/N	ase	mean	sd
0.7	0.71	0.68	0.33	2.5	0.009	4.7	0.9

lower alpha upper 95% confidence boundaries
0.69 0.7 0.72

Reliability if an item is dropped:

	raw_alpha	std.alpha	G6(smc)	average_r	S/N	alpha	se
A1-	0.72	0.73	0.67	0.40	2.6	0.0087	
A2	0.62	0.63	0.58	0.29	1.7	0.0119	
A3	0.60	0.61	0.56	0.28	1.6	0.0124	
A4	0.69	0.69	0.65	0.36	2.3	0.0098	
A5	0.64	0.66	0.61	0.33	1.9	0.0111	

1

Warning message:

```
In alpha(bfi[1:5], check.keys = TRUE) :
```

Some items were negatively correlated with total score and were ^{12/17/8}

R functions will return objects without necessarily telling you

1. The basic logic of R is that you can do lots of calculations, but you might not want all the output.
 2. The output is there, to be processed by other functions if you want, but you probably don't want to see all of it unless you ask.,
 3. Thus, alpha returns the scores based upon the scales you asked for, but doesn't show them, because they are so many,
 4. The `str` command tells you the structure of an object. The names will just list the names of the objects.

names and str of alpha output

R code

```
names(scores)  
str(scores)
```

```

names(scores)
[1] "total"           "alpha.drop"      "item.stats"      "response.fre
"scores"            "nvar"          "boot.ci"
[9] "boot"             "Unidim"         "Fit"            "call"

$ total           :'data.frame':   1 obs. of  8 variables:
..$ raw_alpha: num  0.703
..$ std.alpha: num  0.713
..$ G6(smc)   : num  0.683
..$ average_r: num  0.332
..$ S/N       : num  2.48
..$ ase       : num  0.00895
..$ mean      : num  4.65
..$ sd        : num  0.898
$ alpha.drop    :'data.frame':   5 obs. of  6 variables:
..$ raw_alpha: num [1:5] 0.719 0.617 0.6 0.686 0.643
..$ std.alpha: num [1:5] 0.726 0.626 0.613 0.694 0.656
..$ G6(smc)   : num [1:5] 0.673 0.579 0.558 0.65 0.605
..$ average_r: num [1:5] 0.398 0.295 0.284 0.361 0.322
..$ S/N       : num [1:5] 2.64 1.67 1.58 2.26 1.9
..$ alpha se  : num [1:5] 0.00873 0.0119 0.01244 0.00983 0.01115
$ item.stats   :'data.frame':   5 obs. of  7 variables:

```

One of the objects of alpha is the scores object

R code

```
describe(scores$scores)
```

But, since there scores for all subjects, but just one score, this is not very interesting.

```

describe(scores$scores)
    vars     n mean   sd median trimmed   mad min max range skew kurt
X1      1 2800 4.65 0.9     4.8     4.73 0.89    1    6     5 -0.76
>

```

Note that alpha has the option of doing cumulative scores (adding up items, or scoring in the unit of the items (the default).

R code

```
scores <- alpha(bfi[1:5], check.keys=TRUE, cumulative=TRUE)
#set the cumulative option to be true
describe(scores$scores)
```

```
describe(scores$scores)
```

vars	n	mean	sd	median	trimmed	mad	min	max	range	skew	kurtosis
x1	1	2800	23.08	4.54	24	23.43	4.45	5	30	25	-0.73

α , $\omega_{hierarchical}$ and β as alternative measures of internal consistency

1. α as the mean split half reliability
 - alpha to find α
 - splitHalf to find all (if $n \leq 16$) or 10,000 random possible split half reliabilities ($n > 16$)
 2. $\omega_{hierarchical}$ and ω_{total} as factor based reliabilities
 - $\omega_{hierarchical}$ estimates general factor saturation
 - Found using omega and omegaSem
 3. β as worst split half reliability as an alternative estimate of the general factor saturation.
 - Found using a hierarchical clustering algorithm (iclust).
 - iclust is also useful for scale construction.

α from alpha and all split halves found using splitHalf

Find α and all split half reliabilities of 5 Agreeableness items and 5 Conscientiousness items from the `bfi` data set included in *psych*.

R code

```
alpha(bfi[1:10]) #find alpha, let it automatically reverse items  
splitHalf(bfi[1:10], keys=c(1,9,10)) #reverse 3 items
```

Reliability analysis

Call: alpha(x = bfi[1:10])

raw_alpha std.alpha G6(smc) average_r S/N ase mean sd

lower alpha upper 95% confidence boundaries

lower alpha a

Split half reliabilities

```
Call: splitHalf(r = bfj[1:10], keys = c(1, 9, 10))
```

Maximum split-half reliability (lambda 4) = 0.81

Cuttman lambda .6 = 0.76

Average split half reliability = 0.73

Guttman lambda 3 (alpha) = 0.74

Minimum split half reliability (beta) = 0.41

All possible split halves of 5 agreeableness and 5 conscientiousness items. Note the one worst one! This is not one construct.

splithalves.pdf

Using the `omega` function

R code

omega(ability, 4)

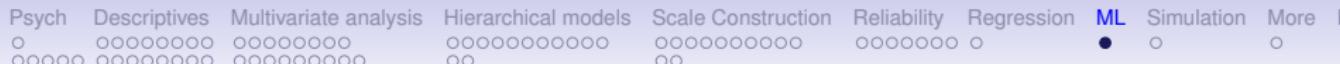
```
Omega
Call: omega(m = ability, nfactors = 4)
Alpha:          0.83
G.6:           0.84
Omega Hierarchical: 0.65
Omega H asymptotic: 0.76
Omega Total      0.86
```

Schmid Leiman Factor loadings greater than .0.2

	<i>g</i>	<i>F1*</i>	<i>F2*</i>	<i>F3*</i>	<i>F4*</i>	<i>h</i> ₂	<i>u</i> ₂	<i>p</i> ₂
reason.4	0.50			0.27		0.34	0.66	0.73
reason.16	0.42			0.21		0.23	0.77	0.76
reason.17	0.55			0.47		0.52	0.48	0.57
reason.19	0.44			0.21		0.25	0.75	0.77
letter.7	0.52		0.35			0.39	0.61	0.69
letter.33	0.46		0.30			0.31	0.69	0.70
letter.34	0.54		0.38			0.43	0.57	0.67
letter.58	0.47		0.20			0.28	0.72	0.78
matrix.45	0.40				0.66	0.59	0.41	0.27
matrix.46	0.40				0.26	0.24	0.76	0.65
matrix.47	0.42					0.23	0.77	0.79
matrix.55	0.28					0.12	0.88	0.65
rotate.3	0.36	0.61				0.50	0.50	0.26
rotate.4	0.41	0.61				0.54	0.46	0.31
rotate.6	0.40	0.49				0.41	0.59	0.39
rotate.8	0.32	0.53				0.40	0.60	0.26

With eigenvalues of:

g F1* F2* F3* F4*



Regression from correlation matrices

1.

Scale construction through “Machine Learning”

1. Supervised Learning was called item analysis in 1930
2. Need to cross validate

Making up data

1.

Miscellaneous functions that are useful

vJoin Merge two files by rownames and column names

scrub Clean up data

`df2latex` One of several functions to create L^AT_EXtables.

`read.file` and `read.clipboard` for convenient input

Athenstaedt, U. (2003). On the content and structure of the gender role self-concept: Including gender-stereotypical behaviors in addition to traits. *Psychology of Women Quarterly*, 27(4), 309–318.

Cohen, J. (1988). *Statistical power analysis for the behavioral sciences* (2nd ed ed.). Hillsdale, N.J.: L. Erlbaum Associates.

Condon, D. M. (2018). *The SAPA Personality Inventory*: An empirically-derived, hierarchically-organized self-report personality assessment model. PsyArXiv /sc4p9/.

Condon, D. M. & Revelle, W. (2014). The International Cognitive Ability Resource: Development and initial validation of a public-domain measure. *Intelligence*, 43, 52–64.

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